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			EPO; JPO;	ĺ
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		US-6020197-\$ or US-6045807-\$ or		
		US-6071889-\$ or US-6197585-\$ or		
		US-6265175-\$ or US-6294346-\$ or		
		US-5411883-\$ or US-6284539-\$).did. or		
		(US-20020009743-\$ or US-20020039789-\$ or		
		US-20020006660-\$).did. or (WO-9949014-\$ or		
		WO-200009669-\$).did.		
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			DERWENT	
_	268	mesencephalon and (neuron\$10 or neural\$5)	USPAT;	2002/09/17 14:34
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_	252	1 '	USPAT;	2002/09/17 14:34
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			EPO; JPO;	
	_	maganashalan and CAU	DERWENT	2002/00/17 11 25
-	2	mesencephalon and SAH	USPAT;	2002/09/17 14:36
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			EPO; JPO;	
1	110	//	DERWENT	0000/00/15 11 11
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		mesencephalon.clm.	EPO; JPO;	
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PCT

WORLD INTELLECTUAL PROPERTY ORGANIZATION International Bureau



INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)-

(51) International Patent Classification 7: WO 00/09669 (11) International Publication Number: C12N 5/10, A61K 48/00 A1 (43) International Publication Date: 24 February 2000 (24.02.00) (21) International Application Number: PCT/US99/18403 (81) Designated States: AU, CA, JP, European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, (22) International Filing Date: 12 August 1999 (12.08.99) NL, PT, SE). Published (30) Priority Data: 09/134,771 12 August 1998 (12.08.98) With international search report. (71) Applicant: SIGNAL PHARMACEUTICALS, INC. [US/US]; 5555 Oberlin Drive, San Diego, CA 92121 (US). (72) Inventors: SAH, Dinah, W.; 8492 Cliffridge Avenue, La Jolla, CA 92037 (US). RAYMON, Heather, K.; 535 Genter Street, La Jolia, CA 92037 (US). (74) Agents: MAKI, David, J. et al.; Seed and Berry LLP, 6300 Columbia, 701 Fifth Avenue, Seattle, WA 98104-7092 (US).

(54) Title: HUMAN MESENCEPHALON CELL LINES AND METHODS OF USE THEREFOR

(57) Abstract

Conditionally-immortalized human mesencephalon cell lines are provided. Such cell lines, which may be clonal, may be used to generate neurons, including dopaminergic neurons. The cell lines and/or differentiated cells may be used for the development of therapeutic agents to prevent and treat a variety of neurological diseases such as Parkinson's disease. The cell lines and/or differentiated cells may also be used in assays and for the general study of mesencephalon cell development and differentiation.

(FILE 'HOME' ENTERED AT 16:14:55 ON 17 SEP 2002)

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     1996:123431 CAPLUS
AN
     124:227790
DN
     Differentiation of the immortalized adult neuronal progenitor
ΤI
     cell line HC2S2 into neurons by regulatable suppression of the
     v-myc oncogene
SO
     Proceedings of the National Academy of Sciences of the United States of
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CODEN: PNASA6; ISSN: 0027-8424
ΑU
     Hoshimaru, Minoru; Ray, Jasodhara; Sah, Dinah W. Y.; Gage, Fred
AB
     A regulatable retroviral vector in which the v-myc oncogene is driven by a
     tetracycline-controlled transactivator and a human cytomegalovirus minimal
     promoter fused to a tet operator sequence was used for conditional
     immortalization of adult rat neuronal progenitor cells. A
     single clone, HC2S2, was isolated and characterized. Two days after the
     addn. of tetracycline, the HC2S2 cells stopped proliferating, began to
     extend neurites, and expressed the neuronal markers
     tau, NeuN, neurofilament 200 kDa, and glutamic acid
     decarboxylase in accordance with the reduced prodn. of the v-myc
     oncoprotein. Differentiated HC2S2 cells expressed large sodium and
     calcium currents and could fire regenerative action potentials. These
     results suggest that the suppression of the v-myc oncogene may be
     sufficient to make proliferating cells exit from cell cycles and induce
     terminal differentiation. The HC2S2 cells will be valuable for studying
     the differentiation process of neurons.
L25 ANSWER 10 OF 21 CAPLUS COPYRIGHT 2002 ACS
     1997:371172 CAPLUS
AN
     127:93276
DN
TT
     Bipotent progenitor cell lines from the human CNS
     Nature Biotechnology (1997), 15(6), 574-580
SO
     CODEN: NABIF9; ISSN: 1087-0156
ΑIJ
     Sah, Dinah W. Y.; Ray, Jasodhara; Gage, Fred H.
AB
     Human central nervous system (CNS) cell lines would substantially
     facilitate drug discovery and basic research by providing a readily renewable source of human {\tt neurons}. We isolated clonal human CNS
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cell lines that had been immortalized with a tetracycline (Tc)-responsive v-myc oncogene; addn. of Tc to the growth medium suppressed the oncoprotein rapidly and virtually completely, allowing differentiation to proceed. Two classes of bipotent precursor cells were immortalized: the first class had a default differentiation pathway of neurons only, and the second class had a default differentiation pathway of neurons and astrocytes. We found that after exposure to different external signals in vitro, the environment is capable of redirecting the fate of a particular cell, even in the case of the bipotent precursor cell whose default differentiation pathway was neurons only. These data suggest that extrinsic cues can prevail over intrinsic determinants in directing cell fate in the human CNS.

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L25 ANSWER 12 OF 21 CAPLUS COPYRIGHT 2002 ACS
     1998:176007 CAPLUS
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     128:229000
TI
     Development of human CNS cell lines and use to study CNS cell development,
     death, and abnormalities
so
     PCT Int. Appl., 76 pp.
     CODEN: PIXXD2
IN
     Sah, Dinah W. Y.; Gage, Fred H.; Ray, Jasodhara
     Conditionally-immortalized human CNS progenitor cell lines are provided.
AB
     Such cell lines, which may be clonal, may be used to generate neurons and/or astrocytes. Such cell lines and/or differentiated
     cells may be used for the development of therapeutic agents to prevent and
     treat a variety of CNS-related diseases. The cell lines are produced by
     transfecting CNS progenitor cells with oncogenes and growing the cells on
     polyornithine/laminin, polylysine/laminin, or fibronectin-treated surfaces
     in culture medium supplemented with proliferation-enhancing factors.
     Suitable oncogenes include v-myc, N-myc, c-myc, p53, SV40 large T antigen, polyoma large T antigen, E1a adenovirus, and the human papillomavirus E7
     protein gene. Such cell lines and/or differentiated cells may also be
     used in assays and for the general study of CNS cell development, death
     and abnormalities. Examples of abnormalities include Alzheimer's disease,
     stroke, traumatic head injuries, and amyotrophic lateral sclerosis.
     PATENT NO.
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AN
     1999:672982 CAPLUS
DN
     131:269274
TI
     PNS cell lines and methods of use therefor
SO
     PCT Int. Appl., 84 pp.
     CODEN: PIXXD2
IN
     Sah, Dinah W. Y.; Raymon, Heather K.
AB
     Conditionally-immortalized PNS progenitor cell lines are provided. Such
     cell lines, which may be clonal, may be used to generate neurons
        The cell lines and/or differentiated cells may be used for the
     development of therapeutic agents to prevent and treat a variety of
     PNS-related diseases. The cell lines and/or differentiated cells may also
     be used in assays and for the general study of PNS cell development, death
     and abnormalities.
     PATENT NO.
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     1999:577009 CAPLUS
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     131:195449
     Conditionally-immortalized human spinal cord cell lines and uses thereof
     in the diagnosis, treatment, and prevention of spinal cord-related
     diseases and injuries
     PCT Int. Appl., 37 pp.
SO
     CODEN: PIXXD2
     Li, Ronghao; Sah, Dinah W. Y.
TN
     Conditionally-immortalized human spinal cord cell lines are provided. In
     one aspect, the present invention provides methods for producing a
     conditionally-immortalized human spinal cord neural precursor
     cell, comprising the steps of: (a) transfecting human spinal cord cells,
     plated on a first surface and in a first growth medium that permit
     proliferation, with DNA encoding a selectable marker and an externally
     regulative growth-promoting gene; and (b) selecting the transfected cells
     on a second surface and in a second growth medium that permit attachment
     and proliferation, and therefrom producing said immortalized cell. In
     certain embodiments, the growth-promoting gene may be an oncogene, such as
     v-myc, and expression of the growth-promoting gene may, but need not, be
     inhibited by tetracycline. Such cell lines, which may be clonal, may be
     used to generate neurons, including motor neurons.
     The cell lines and/or differentiated cells may be used for the development
     of therapeutic agents to prevent and treat a variety of spinal
     cord-related diseases and injuries. The cell lines and/or differentiated
     cells may also be used in assays and for the general study of spinal cord
     cell development and differentiation.
     PATENT NO.
                      KIND DATE
                                            APPLICATION NO. DATE
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     WO 9945103
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L2
             10 DUP REM L1 (13 DUPLICATES REMOVED)
             10 SORT L2 PY
L3
L4
          31741 S MESENCEPHALON
L5
            208 S L4 AND (NEUR? (S) PROGENITOR)
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L7
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              5 SORT L7 PY
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L10
L11
            141 DUP REM L10 (299 DUPLICATES REMOVED)
L12
            141 FOCUS L11 1-
              5 S L11 AND HC2S2
L13
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L13 ANSWER 5 OF 5 CAPLUS COPYRIGHT 2002 ACS
AN
     1997:568304 CAPLUS
DN
     127:201026
     Regulatable retroviral vector containing v-myc oncogene for
TΤ
     immortalization of adult neuronal progenitor
     cells
SO
     PCT Int. Appl., 42 pp.
     CODEN: PIXXD2
IN
     Gage, Fred H.; Ray, Jasodhara; Hoshimaru, Minoru
     A novel regulatable retroviral vector in which the v-myc oncogene is
AB
     driven by a tetracycline-controlled transactivator and a human
     cytomegalovirus minimal promoter fused to tet operator sequence useful for
     immortalization of adult neuronal progenitor
     cells is provided. Producer cell lines which produce high titers of the
     recombinant retrovirus are also provided. This general method is
     exemplified by the retroviral vector LINXv-myc. HC2S2 cells
     from adult rat hippocampus were infected with the retroviral vectors.
     HC2S2 cells, derived from an immortalized
     neuronal progenitor cell, were differentiated into
     neurons after suppression of the v-myc oncogene.
     PATENT NO.
                     KIND DATE
                                           APPLICATION NO. DATE
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ΤI
     Differentiation of the immortalized adult neuronal
     progenitor cell line HC2S2 into neurons by regulatable
     suppression of the v-myc oncogene.
SO
     PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF THE UNITED STATES OF
     AMERICA, (1996 Feb 20) 93 (4) 1518-23.
     Journal code: 7505876. ISSN: 0027-8424.
ΑIJ
    Hoshimaru M; Ray J; Sah D W; Gage F H
     A regulatable retroviral vector in which the v-myc oncogene is driven by a
     tetracycline-controlled transactivator and a human cytomegalovirus minimal
```

promoter fused to a tet operator sequence was used for conditional

immortalization of adult rat neuronal progenitor
cells. A single clone, HC2S2, was isolated and characterized. Two days
after the addition of tetracycline, the HC2S2 cells stopped proliferating,
began to extend neurites, and expressed the neuronal
markers tau, NeuN, neurofilament 200 kDa, and glutamic acid
decarboxylase in accordance with the reduced production of the v-myc
oncoprotein. Differentiated HC2S2 cells expressed large sodium and calcium
currents and could fire regenerative action potentials. These results
suggest that the suppression of the v-myc oncogene may be sufficient to
make proliferating cells exit from cell cycles and induce terminal
differentiation. The HC2S2 cells will be valuable for studying the
differentiation process of neurons.

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make proliferating cells exit from cell cycles and induce terminal
     differentiation. The HC2S2 cells will be valuable for studying the
     differentiation process of neurons.
L12 ANSWER 1 OF 141 CAPLUS COPYRIGHT 2002 ACS
     1997:568304 CAPLUS
     Regulatable retroviral vector containing v-myc oncogene for
TΤ
     immortalization of adult neuronal progenitor
SO
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     CODEN: PIXXD2
     Gage, Fred H.; Ray, Jasodhara; Hoshimaru, Minoru
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     A novel regulatable retroviral vector in which the v-myc oncogene is
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     driven by a tetracycline-controlled transactivator and a human
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     immortalization of adult neuronal progenitor
     cells is provided. Producer cell lines which produce high titers of the
     recombinant retrovirus are also provided. This general method is
     exemplified by the retroviral vector LINXv-myc. HC2S2 cells from adult
     rat hippocampus were infected with the retroviral vectors. HC2S2 cells,
     derived from an immortalized neuronal
     progenitor cell, were differentiated into neurons after
     suppression of the v-myc oncogene.
                     KIND DATE
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    AU 721727
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    EP 892851
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            AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT,
             IE, FI
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L12 ANSWER 2 OF 141 CAPLUS COPYRIGHT 2002 ACS
ΑN
    1999:20170 CAPLUS
     130:245944
TI
    The use of neural progenitor cells for therapy in the
     CNS disorders
     CNS Regeneration (1999), 183-201. Editor(s): Tuszynski, Mark H.;
     Kordower, Jeffrey H. Publisher: Academic, San Diego, Calif.
     CODEN: 67CYA3
ΑU
     Ray, Jasodhara; Palmer, Theo D.; Shihabuddin, Lamya S.; Gage, Fred H.
AB
     A review with 84 refs. In recent years a significant no. of neurol.
     diseases have been defined at the mol. level. Somatic gene therapy using
     genetically modified non-neuronal cells expressing therapeutic factors
    have been successfully used in animal models of neurodegenerative
     diseases. Ability to grow central nervous system (CNS)-derived
    neural progenitor cells has proven to be extremely
     useful to study a diverse phenomenon including the fate choice,
     differentiation, and synaptic maturation of cells. Immortal or
    perpetual cultures of neural progenitor cells
     implanted into the rodent brain survive, migrate, and integrate in the
    host cytoarchitecture. These cells can be genetically modified to express
     therapeutic gene products. The ability of the implanted cells to
     integrate in the host brain and express transgene products in situ offer
    potential approaches for gene therapy in certain CNS diseases. The
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utility of this approach has already been explored in animal models of neurodegenerative diseases. This chapter reviews the recent advances made in understanding the nature and potentiality of neural progenitor cells in vitro and in vivo as well as their possible use for cell replacement and gene therapy.

L12 ANSWER 5 OF 141 MEDLINE

AN 95212010 MEDLINE

TI A short term analysis of the behaviour of conditionally immortalized neuronal progenitors and primary neuroepithelial cells implanted into the fetal rat brain.

SO BRAIN RESEARCH. DEVELOPMENTAL BRAIN RESEARCH, (1994 Dec 16) 83 (2) 197-208.

Journal code: 8908639. ISSN: 0165-3806.

AU Cattaneo E; Magrassi L; Butti G; Santi L; Giavazzi A; Pezzotta S

Conditionally immortalized (temperature-sensitive) striatal-derived neuronal progenitor cell lines and primary neuroepithelial cells were transplanted into the CNS of gestational day 15-16 rat fetuses using an 'in utero' surgical procedure. Each fetus received 2.5-3 x 10(4) donor cells previously labelled in vitro by incubation with 5-bromo-2'-deoxyuridine (BrdU). At 5 days following transplantation, 69% of the fetuses were still alive. Engrafted cells were detected by BrdU immunohistochemistry, and the appearance of the engrafted cells and the time course of Nestin and PCNA expression were measured at 6, 24, 64 h and 5 days after transplantation. The evolution of Large T-Antigen immunoreactivity in engrafted temperature-sensitive (ts) cells was also evaluated at the above time intervals. The results indicate that the majority of the implanted cells were aggregated into clusters 24 h after transplantation. These clusters were not visible at 6 h, when most of the cells were isolated. The clusters were located in both the ventricles and parenchyma. These findings were common to both ts cells and striatal primary neuroepithelial cells. At 64 h and 5 days, isolated cells associated with the germinal layer and scattered throughout the parenchyma were also found. In the clusters, Nestin expression decreased proportionally with time following transplantation. Furthermore, Large T-Antigen immunoreactivity disappeared from ts cells between 6 and 24 h after transplantation. Finally, measurements of the temporal evolution of PCNA expression within the clusters indicate a progressive reduction in the mitotic activity of the transplanted cells. The results demonstrate that striatal primary neuroepithelial cells and conditionally immortalized neuronal

progenitors can survive, migrate and/or compartimentalize into clusters whilst changing their antigenic properties and ability to proliferate.

L12 ANSWER 7 OF 141 MEDLINE

AN 1998063822 MEDLINE

ΑIJ

TI In vitro pattern formation during neurogenesis in neuroectodermal progenitor cells immortalized by p53-deficiency.

SO INTERNATIONAL JOURNAL OF DEVELOPMENTAL NEUROSCIENCE, (1997 Oct) 15 (6) 795-804.

Journal code: 8401784. ISSN: 0736-5748.

Schlett K; Herberth B; Madarasz E

In vitro neural differentiation was induced in a p53-deficient immortalized neuroectodermal progenitor cell line, NE-4C, by treatment with retinoic acid [K. Schlett and E. Madarasz (1997) J. Neurosci. Res. 47, 405-416]. Rearrangement of nestin filaments was an early marker of neuron-formation. The increase in neurofilament protein content was accompanied by a decrease in the expression of nestin filaments in induced precursors. Cells with astroglial features appeared with a delay of 4-5 days compared to the appearence of neurons. Future neurons were sorted out from the substrate-attached population of apparently non-induced cells. The sorting out of future neurons resembled the separation of neural precursors in vivo. The continuous changes in the shape and also in the position of the cells resulted in the formation of characteristic morphological patterns. On the basis of morphological changes, five characteristic stages of in vitro neural differentiation were distinguished. The analysis of the morphological

changes revealed that cell-to-cell interactions played an essential role in the cell fate decision made by induced precursors. Our observations indicate that the NE-4C cell line can serve as an in vitro model to investigate some early steps of neurogenesis.

- L12 ANSWER 10 OF 141 CAPLUS COPYRIGHT 2002 ACS
- AN 1994:266389 CAPLUS
- DN 120:266389
- TI Immortalization of neural cells with the c-myc and N-myc proto-oncogenes
- SO NeuroProtocols (1993), 3(3), 200-13 CODEN: NEPREV; ISSN: 1058-6741
- AU Bernard, Ora
- AB A review, with 59 refs. The c-myc and the N-myc proto-oncogenes were employed to immortalize neural progenitor cells. Infection of neural precursors isolated from the mouse at the 10th day of embryonic development (E10) with myc-contg. retroviruses resulted in immortalized cell lines representing bipotential E10 neuroepithelial cells. These cell lines have the capacity to differentiate into both glial and neuronal cells either spontaneously in the case of the Zen(myc) cell lines or after addn. of fibroblast growth factor to the Dol(myc) cell lines. Infection of migrating neural crest cells with the myc retroviruses gave rise to three different types of immortalized cell lines: (i) cell lines resembling freshly isolated neural crest cells; (ii) cell lines that can differentiate into cells expressing Schwann cell markers when grown at high cell concns.; and (iii) cell lines that have the ability to differentiate in culture to process-bearing cells which expressed neuronal markers or have the characteristics of Schwann cells. Olfactory epithelial cell lines were generated by infection with Zen retrovirus bearing the N-myc proto-oncogene. Some of the cell lines resemble basal cells and others grow as bipolar cells resembling neurons and expressing the neuronal marker neurofilaments.
- L12 ANSWER 13 OF 141 BIOSIS COPYRIGHT 2002 BIOLOGICAL ABSTRACTS INC.
- AN 2001:519851 BIOSIS
- TI Growth factor addition influences immortalized neural progenitor cell proliferation and differentiation.
- SO Society for Neuroscience Abstracts, (2001) Vol. 27, No. 1, pp. 941. print. Meeting Info.: 31st Annual Meeting of the Society for Neuroscience San Diego, California, USA November 10-15, 2001 ISSN: 0190-5295.
- AU Teppen, T. L. (1); Peterson, D. A. (1)
- Neural stem cells provide a potential therapy to repair neuronal loss experienced with neurodegenerative disease. C17-2 neural progenitor cells (provided by E.Y. Snyder) are reported to display lower mitotic activity when cultured in completely defined media (N2) compared to serum supplementation. Addition of specific growth factors has been reported to enhance proliferation. To evaluate the effect of the growth factors bFGF and BDNF on proliferation and differentiation, C17-2 cells were cultured in defined (N2) media alone or supplemented with bFGF, BDNF or bFGF+BDNF in the absence of serum. Cell number and phenotype were compared with cells maintained in serum supplemented media. Cells cultured in defined medium supplemented with bFGF, BDNF or both factors together displayed a change in morphology compared with cells cultured in serum. While serum supplementation produced a cell monolayer, the addition of growth factors resulted in the formation of cell clusters appearing as proliferative cell colonies. A higher level of proliferation was observed across the bFGF and bFGF+BDNF conditions than with BDNF alone. Previous findings that bFGF and BDNF display a synergistic interaction were not observed. All defined media conditions appeared to have a lower growth rate (between 39-65%) than that of serum containing cultures. Results suggest that both bFGF and BDNF are mitogens for C17-2 cells. We are currently assessing the effect of growth factors on neuronal differentiation. These findings suggest C17-2 cells have the potential to proliferate and differentiate in response to specific environmental manipulations.
- L12 ANSWER 14 OF 141 MEDLINE
- AN 1998031251 MEDLINE

- TI Immortalized neural progenitor cells for CNS gene transfer and repair.
- SO TRENDS IN NEUROSCIENCES, (1997 Nov) 20 (11) 530-8. Ref: 58 Journal code: 7808616. ISSN: 0166-2236.
- AU Martinez-Serrano A; Bjorklund A
- Immortalized multipotent neural stem and AB progenitor cells have emerged as a highly convenient source of tissue for genetic manipulation and ex vivo gene transfer to the CNS. Recent studies show that these cells, which can be maintained and genetically transduced as cell lines in culture, can survive, integrate and differentiate into both neurons and glia after transplantation to the intact or damaged brain. Progenitors engineered to secrete trophic factors, or to produce neurotransmitter-related or metabolic enzymes can be made to repopulate diseased or injured brain areas, thus providing a new potential therapeutic tool for the blockade of neurodegenerative processes and reversal of behavioural deficits in animal models of neurodegenerative diseases. With further technical improvements, the use of immortalized neural progenitors may bring us closer to the challenging goal of targeted and effective CNS repair.
- L12 ANSWER 16 OF 141 MEDLINE
- AN 97084035 MEDLINE
- TI Conditionally immortalized neural progenitor cell lines integrate and differentiate after grafting to the adult rat striatum. A combined autoradiographic and electron microscopic study.
- SO BRAIN RESEARCH, (1996 Oct 21) 737 (1-2) 295-300. Journal code: 0045503. ISSN: 0006-8993.
- AU Lundberg C; Field P M; Ajayi Y O; Raisman G; Bjorklund A
- Neural progenitor cell lines, generated by conditional AΒ immortalization from the embryonic CNS, have previously been shown to survive and integrate after transplantation to the adult brain. The present study was designed to investigate the in vivo differentiation and morphological features of grafted neural progenitors using combined autoradiography and transmission electron microscopy of two temperature-sensitive neural progenitor cell lines, HiB5 and ST14A, labeled with 3H-thymidine prior to grafting. Two weeks after transplantation to the striatum the cells were found dispersed over an area extending about 1.5 mm from the injection site. Labeled cells located within the myelinated fiber bundles of the internal capsule were closely associated with myelinated axons and presented profiles similar to oligodendrocytes, while most of the grafted cells in the grey matter had morphological features of astroglia. Some labeled cells occurred also in close association with small blood vessels, morphologically resembling host pericytes. The results show that the immortalized neural progenitors can differentiate into mature glial cells, including astrocytes, oligodendrocytes and pericytes, after implantation into the adult striatum. The ability of the cells to become fully integrated with the resident glial population suggests that they will be highly useful as vehicles for intracerebral transgene expression in ex vivo gene transfer.
- L12 ANSWER 17 OF 141 MEDLINE
- AN 94376081 MEDLINE
- TI FGF and EGF are mitogens for immortalized neural progenitors.
- SO JOURNAL OF NEUROBIOLOGY, (1994 Jul) 25 (7) 797-807.
- Journal code: 0213640. ISSN: 0022-3034. AU Kitchens D L; Snyder E Y; Gottlieb D I
- AB Individual neural progenitors, derived from the external germinal layer of neonatal murine cerebellum, were previously immortalized by the retrovirus-mediated transduction of avian myc (v-myc). C17-2 is one of those clonal multipotent progenitor cell lines (Snyder et al., 1992, Cell 68: 33-51; Ryder et al., 1990, J. Neurobiol. 21:356-375). When transplanted into newborn mouse cerebellum (CB), the cells participate in normal CB development; they engraft in a cytoarchitecturally appropriate, nontumorigenic manner and differentiate into multiple CB cell types (neuronal and glial) similar to endogenous progenitors (Snyder et al., 1992, as

above). They also appear to engraft and participate in the development of multiple other structures along the neural axis and at multiple other stages (Snyder et al., 1993, Soc. Neurosci. Abstr. 19). Thus conclusions regarding these immortalized progenitors may be applicable to endogenous neural progenitors in vivo. To help identify and analyze factors that promote differentiation of endogenous progenitors, we first investigated the ability to maintain C17-2 cells in a defined, serum-free medium (N2). The cells survive in vitro in N2 but undergo mitosis at a very low rate. Addition of epidermal growth factor (EGF), however, either from mouse submaxillary gland or the human recombinant protein, appreciably stimulates thymidine incorporation and cell division approximately threefold. Basic fibroblast growth factor (bFGF) is an even more potent mitogen, promoting thymidine incorporation, cell division, and a net increase in cell number equal to that in serum. Both EGF and bFGF are active at very low nanomolar concentrations, suggesting that they interact with their respective receptors rather than a homologous receptor system. The findings demonstrate that C17-2 cells can be maintained and propagated in a fully defined medium, providing the basis for analysis of other growth and differentiation factors. That EGF and particularly bFGF are mitogenic for these cells is in accord with recent observations on primary neural tissue (Reynolds and Weiss, 1992, Science 255:1707-1710; Kilpatrick and Bartlett, 1993, Neuron 10:255-265; Ray et al., 1993, Proc. Natl. Acad. Sci. USA 90:3602-3606) suggesting that bFGF and EGF responsiveness may be fundamental properties of neural progenitors.

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